

Proficiency Test Instructions for Participants

15 & 16 MAY 2017 RUN

PURPOSE

To provide for detailed instructions for participants in compliance with the requirements of ISO 17043, clause 4.6.1

INSTRUCTIONS

Scheme reference	Samples			Samples reconstitution	Testing timeframe	Approved test methods	Sheet to be used for submission of results to JGK	Unit of measurement (Results)	Number of test replicates per sample
	Type	Identification (as per the labels)	Storage						
JGKPT01 Antimicrobial susceptibility	Animal sp.: <u>Cat</u> 2 FD cultures (clinical pathogens)	ATB01C – ATB02C	2-8°C	Add 1 ml sterile DH ₂ O to the vial	Test immediately after reconstitution Observe deadline for submission of results	Disc diffusion technique (Kirby Bauer)	R.E.F 1.2 (supplied)	Zone Ø in millimeters (express results: S, I, R) 'S': sensitive 'I': intermediate 'R': resistance	Culture: 1 rep. Susceptibility: Supplied discs: 3 repl. Own discs: 1 rep.
	Antimicrobial discs (x18)	Tetracycline 30 µg (x6) Ciprofloxacin 5 µg (x6) Erythromycin 15 µg (x6)		Discs are ready to use (Commercial discs)					
JGKPT03 Brucella: RBT	6 FD bovine sera	RBT01 – RBT06	2-8°C	Add 250 µl sterile DH ₂ O to the vial	Test within 5 days after reconstitution Observe deadline for submission of results	RBT	R.E.F 3.5 (supplied)	Positive/Negative	1 rep.
JGKPT04 Brucella: CFT	6 FD bovine sera	CFT01 – CFT06	2-8°C	Add 250 µl sterile DH ₂ O to the vial	Test within 5 days after reconstitution Observe deadline for submission of results	CFT (DAFF approved SOP for SA Labs)	R.E.F 3.5 (supplied)	CFTIU/ml	2 rep.
JGKPT05 Brucella: MRT	3 Preserved cow's milk	CAM01 – CAM03	2-8°C	N/A Ready to process	Preferably immediately but no later than 5 days from date of dispatch	MRT	R.E.F 3.5 (supplied)	Positive/Negative	1 rep.
JGKPT06 Brucella: Culture & Identification	3 FD cultures	CAC01 – CAC03	2-8°C	Add 1 ml sterile DH ₂ O to the vial	Test immediately after reconstitution Observe deadline for submission of results	Conventional microbiology and Molecular technique	R.E.F 06 (supplied)	Presence/Absence: <u>Brucella abortus</u> Strain: <u>Field or vaccine</u>	1 rep.

JGKPT07 Brucella: Slides stain & ID	4 unstained slides (4 replicates each)	Slide 1 – slide 4	Room T° (dust free)	N/A	Anytime, as long as deadline for submission of results is observed	Appropriate staining techniques	R.E.F 06 (supplied)	Positive/Negative <u>Brucella organisms</u>	1 rep.
JGKPT08 <i>Avibacterium paragallinarum</i>	3 FD cultures	AVI01 – AVI03	2-8°C	If FD form: Add 1 ml sterile DH ₂ O to the vial If Liquid form: N/A, ready to process	FD form: Test within 5 days after reconstitution Liquid form: Test within 2 days of receipt	Standard veterinary microbiology	R.E.F 08-17 (supplied)	Positive/Negative <u>Avibacterium paragallinarum</u>	1 rep.
JGKPT12 Salmonella in Poultry: Culture & Identification	3 FD cultures	SAP01 – SAP03	2-8°C	If FD form: Add 1 ml sterile DH ₂ O to the vial If Liquid form: N/A, ready to process	FD form: Test within 5 days after reconstitution Liquid form: Test within 2 days of receipt	Standard veterinary microbiology	R.E.F 08-17 (supplied)	Full ID of the bacteria isolated Target: <i>Salmonella</i>	1 rep.
JGKPT13 Salmonella in Mammals: Culture & Identification	3 FD cultures	SAM01 – SAM03	2-8°C	If FD form: Add 1 ml sterile DH ₂ O to the vial If Liquid form: N/A, ready to process	FD form: Test within 5 days after reconstitution Liquid form: Test within 2 days of receipt	Standard veterinary microbiology	R.E.F 08-17 (supplied)	Full ID of the bacteria isolated Target: <i>Salmonella</i>	1 rep.
JGKPT61-64 Trichomonas and Campylobacter: Culture/PCR	5 Cultures in transport media	SW01 – SW05	2-8°C	N/A Ready to process	Preferably immediately but no later than 5 days after dispatch	Conventional microbiology and Molecular technique	R.E.F 61.4 (supplied)	Presence/Absence: <u>Trichomonas foetus</u> , <u>Campylobacter fetus</u>	1 rep.

Treatment of samples

No special treatment should be given to the PT samples. They [samples] should be handled in the same manner as all the other samples routinely tested in the laboratory.

Handling and safety requirements

Contact surfaces must be decontaminated with 70% alcohol or other suitable disinfectants. Used laboratory utensils must be cleaned and sterilized as per laboratory standard operating procedure. Unused materials and waste must be removed as per laboratory waste management protocol. This involves autoclaving and/or removal by an approved waste removal company.

Specific environmental conditions required for the participant to conduct tests

JGKPT06 (Brucella Culture) must be handled in a biological safety cabinet class 2+

Results recording

Only the supplied excel “Result entry forms” [R.E.F] should be used to capture the PT results. The Participants must fill in the cover page and answer all the questions before capturing the results on R.E.F.

Submission of PT results to JGK

All results must be submitted on or before the **6th of June 2017**. **Any results received after this date will not be included in the analysis.**

Enquiries

For any inquiry, please use the contact details provided above to contact JGK

Return of the proficiency test items

None

FD: freeze-dried