

## Proficiency Test Instructions for Participants

### RUN 4: 16 OCTOBER 2017

#### PURPOSE

To provide for detailed instructions for participants in compliance with the requirements of ISO 17043, clause 4.6.1

#### INSTRUCTIONS

Scheme Description and Code	Samples			Samples reconstitution	Testing timeframe	Approved test methods	Sheet to be used for submission of results to JGK	Unit of measurement (Results)	Number of test replicates per sample
	Type	Identification (as per the labels)	Storage						
Antibiogram – mastitis pathogens <b>JGKPT02</b>	2 cultures (liquid) 18 ATB discs	ATB01 – ATB02 Discs: Penicillin 10, Polymixin B 300, Sulphafurazole 300	2-8°C	Ready to process	Test within 2 days of dispatch Observe deadline for submission of results	Kirby-Bauer Disc diffusion	R.E.F 02 (supplied)	Zone Ø in millimeters (Results: S, I, R)	Culture: 1 <u>Susceptibility:</u> Supplied discs: 3 Own discs: 1
Antimicrobial residues – meat (β-lactams, Tetracycline, Nonspecific) <b>JGKPT32-34</b>	4 pieces of meat (25 g)	RES01F – RES04F	2-8°C	Ready to process	Test within 5 days of dispatch Observe deadline for submission of results	Rapid test Microbiological Chromatography	R.E.F 32-44 (supplied)	Penicillin/Tetracycline OR Presence/Absence	1
Antimicrobial residues – milk (β-lactams, Tetra, Nonspecific) <b>JGKPT29, 30, 31</b>	5 milk (liquid)	RES01M – RES05M	2-8°C	Ready to process	Test within 5 days of dispatch Observe deadline for submission of results	Rapid test Microbiological Chromatography	R.E.F 26-31 (supplied)	Presence/Absence	1
<i>Campylobacter jejuni</i> – meat (Detection) <b>JGKPT35</b>	5 pieces of meat (25 g)	FH01 – FH05	2-8°C	Ready to process	Test within 2 days of dispatch Observe deadline for submission of results	Microbiological (conventional & automated)	R.E.F 32-44 (supplied)	Presence/Absence	1
<i>E. coli</i> O157 – meat (Detection) <b>JGKPT37</b>									
<i>Listeria monocytogenes</i> – meat (Detection) <b>JGKPT42</b>									
<i>Salmonella</i> species – meat <b>JGKPT43</b>									
Mastitis pathogens – milk	2 cultures	MAST01 – MAST02	2-8°C	Ready to process	Test within 2 days of	Culture & ID	R.E.F 59-60	Target bacteria	1

JGKPT60	(liquid)				dispatch Observe deadline for submission of results		(supplied)		
Milk hygiene: Coliforms count JGKPT27	3 milk (Freeze-dried)	MHCOL01 – MHCOL03	2-8°C	Add 10 ml sterile DH <sub>2</sub> O to the vial 2 dilutions (10 fold) before testing	Test immediately after reconstitution Observe deadline for submission of results	3M Petri Films	R.E.F 26-31	Counts	2
Milk hygiene: Total bacteria count JGKPT26	3 milk (Freeze-dried)	MHTBC01 – MHTBC03	2-8°C	Add 10 ml sterile DH <sub>2</sub> O to the vial 2 Dilutions (10 fold) before testing	Test immediately after reconstitution Observe deadline for submission of results	3M Petri Films	R.E.F 26-31 (supplied)	Counts	2
Milk hygiene: <i>E. coli</i> count JGKPT28	3 milk (Freeze-dried)	MHEC01 – MHEC03	2-8°C	Add 10 ml sterile DH <sub>2</sub> O to the vial No dilution: direct plating	Test immediately after reconstitution Observe deadline for submission of results	3M Petri Films	R.E.F 26-31 (supplied)	Counts	2
Somatic cells counts – milk JGKPT59	6 Milk (liquid) (20 ml)	SCC01 – SCC06	2-8°C	Mix thoroughly before testing	Test within 5 days of dispatch Observe deadline for submission of results	Cytology	R.E.F 59-60 (supplied)	Counts	3
Total viable count – meat (Enumeration) JGKPT44	2 FD cultures	TVC01 – TVC02	2-8°C	Add 1ml sterile DH <sub>2</sub> O to the vial Stir and test	Test immediately after reconstitution Observe deadline for submission of results	Microbiology (conventional & automated)	R.E.F 32-44 (supplied)	Counts	2 (averaged)

#### Treatment of samples

No special treatment should be given to the PT samples. They [samples] should be handled in the same manner as all the other samples routinely tested in the laboratory.

#### Handling and safety requirements

Contact surfaces must be decontaminated with 70% alcohol or other suitable disinfectants. Used laboratory utensils must be cleaned and sterilized as per laboratory standard operating procedure. Unused materials and waste must be removed as per laboratory waste management protocol. This involves autoclaving and/or removal by an approved waste removal company.

#### Specific environmental conditions required for the participant to conduct tests

None

#### Results recording

Only the supplied excel “Result entry forms” [R.E.F] should be used to capture the PT results. The Participants must fill in the cover page and answer all questions before capturing the results on R.E.F.

#### Submission of PT results to JGK

All results must be submitted on or before the **6<sup>th</sup> of November 2017**. **Any results received after this date will not be included in the analysis.**

#### Enquiries

For any inquiry, please use the contact details provided above to contact JGK

#### Return of the proficiency test items

None