

Tel: +27 18 294 4528 Fax: +27 86 650 1716 Cell: +27 82 562 1858

Email 1: <u>kangumbaj@gmail.com</u> Email 2: <u>joule@jgklab.co.za</u> Website: www.jgklab.co.za

Proficiency Test Instructions for Participants

04 SEPTEMBER 2017 RUN

PURPOSE

To provide for detailed instructions for participants in compliance with the requirements of ISO 17043, clause 4.6.1

INSTRUCTIONS

Scheme reference	Samples			Samples		Approved test	Sheet to be used for	Unit of moscuroment	Number of test
	Type and number	Identification (as per the labels)	Storage	reconstitution	Testing timeframe	methods	submission of results to JGK	(Results)	replicates per sample
JGKPT12 Salmonella – poultry	3 FD cultures	SAP01 – SAP03	2-8°C	Add 1 ml sterile DH ₂ O to the vial; mix well	Test within 2 days after reconstitution. Keep cool	Clinical veterinary microbiology	R.E.F 12-13 (supplied)	Presence/absence ID of target salmonella	1 rep
JGKPT13 Salmonella – mammals	3 FD cultures	SAM01 – SAM03	2-8°C	Add 1 ml sterile DH ₂ O to the vial; mix well	Test within 2 days after reconstitution Keep cool	Clinical veterinary microbiology	R.E.F 12-13 (supplied)	Presence/absence ID of target salmonella	1 rep
JGKPT24 Pig disease scenario	3 cultures (FD or swabs in transport medium)	BAC01P – BAC03P	2-8°C	<u>FD</u> : add 1 ml sterile DH₂O to the vial; mix well <u>Swabs</u> : ready to use	<u>FD</u> : test within 2 days after reconstitution <u>Swabs</u> : test immediately	Clinical veterinary microbiology	R.E.F 24-25 (supplied)	Full ID of target bacteria responsible for the condition(s) described in scenario	1 rep.
JGKPT25 Sheep/goat disease scenario	3 cultures (FD or swabs in transport medium)	BAC01SG – BAC03SG	2-8°C	<u>FD</u> : add 1 ml sterile DH ₂ O to the vial; mix well <u>Swabs</u> : ready to use	<u>FD</u> : test within 2 days after reconstitution <u>Swabs</u> : test immediately	Clinical veterinary microbiology	R.E.F 24-25 (supplied)	Full ID of target bacteria responsible for the condition(s) described in scenario	1 rep.
JGKPT65 Bovine viral diarrhea – serum	6 bovine sera (liquid)	BVD01 – BVD06	2-8°C	N/A Ready to process as per your SOP	Test within 2 days of dispatch Keep cool	ELISA	R.E.F 65-75 (supplied)	Optical density (OD) and interpretation of results as Pos/Neg	2 rep.
JGKPT67 Fasciola hepatica – serum	6 bovine sera (liquid)	LF01 – LF06	2-8°C	N/A Ready to process as per your SOP	Test within 2 days of dispatch Keep cool	ELISA	R.E.F 65-75 (supplied)	OD and interpretation of results as Pos/Neg	2 rep.
JGKPT69	6 poultry sera	AI01 – AI06	2-8°C	N/A	Test within 2 days of	ELISA	R.E.F 65-75	ELISA: ODs	2 rep.

Page 1 of 3

Document: IFP 01.01/2017

Authorized by Dr. Joule Kangumba

February 2017

Avian influenza – serum	(liquid)			Ready to process as per your SOP	dispatch Keep cool	HA/HI	(supplied)	<u>HA/HI</u> : Titres And: Interpretation of results as Pos/Neg	
JGKPT71 Infectious bronchitis – serum	6 poultry sera (liquid)	IBV01 – IBV06	2-8°C	N/A Ready to process as per your SOP	Test within 2 days of dispatch Keep cool	ELISA	R.E.F 65-75 (supplied)	OD and interpretation of results as Pos/Neg	2 rep.
JGKPT72 Newcastle disease virus – serum	6 poultry sera (liquid)	NDV01 – NDV06	2-8°C	N/A Ready to process as per your SOP	Test within 2 days of dispatch Keep cool	ELISA HA/HI	R.E.F 65-75 (supplied)	<u>ELISA</u> : ODs <u>HA/HI</u> : Titres And: Interpretation of results as Pos/Neg	2 rep.
JGKPT73 Salmonella enteritidis – serum	6 poultry sera (liquid)	SE01 – SE06	2-8°C	N/A Ready to process as per your SOP	Test within 2 days of dispatch Keep cool	RPAT	R.E.F 65-75 (supplied)	Positive/Negative	1 rep.
JGKPT74 Mycoplasma gallisepticum – serum	6 poultry sera (liquid)	MG01 – MG06	2-8°C	N/A Ready to process as per your SOP	Test within 2 days of dispatch Keep cool	RPAT ELISA	R.E.F 65-75 (supplied)	<u>RPAT</u> : Pos/Neg <u>ELISA</u> : OD and interpretation of results as Pos/Neg	RPAT: 1 rep ELISA: 2 rep.
JGKPT76 Avian influenza virus isolation	3 suspensions (frozen)	AIVI01 – AIVI03	Frozen (-80 °C)	Adjust to ambient temperature and process as per SOP	When convenient if stored as directed Observe deadline	Virus isolation in embryonated eggs	R.E.F 76-79 (supplied)	Positive/Negative Virus quantification where applicable	Qualitat: 1 rep. Quantit: 2 rep.
JGKPT77 Avian influenza PCR	3 suspensions (frozen)	AIPCR01 – AIPCR03 (or as indicated on sample vials)	Frozen (-80 °C)	Adjust to ambient temperature and process as per SOP	When convenient if stored as directed Observe deadline	Molecular techniques	R.E.F 76-79 (supplied)	Positive/Negative Or as directed on result sheet	1 rep. Or as directed on result sheet
JGKPT78 Newcastle disease virus isolation	3 suspensions (frozen)	NDVI01 – NDVI03	Frozen (-80 °C)	Adjust to ambient temperature and process as per SOP	When convenient if stored as directed Observe deadline	Virus isolation in embryonated eggs	R.E.F 76-79 (supplied)	Positive/Negative Virus quantification where applicable	Qualitat: 1 rep. Quantit: 2 rep.
JGKPT79 Newcastle disease virus PCR	3 suspensions (frozen)	NDPCR01 – NDPCR03 (or as indicated on sample vials)	Frozen (-80 °C)	Adjust to ambient temperature and process as per SOP	When convenient if stored as directed Observe deadline	Molecular techniques	R.E.F 76-79 (supplied)	Positive/Negative Or as directed on result sheet	1 rep. Or as directed on result sheet

Treatment of samples

No special treatment should be given to the PT samples. They [samples] should be handled in the same manner as all the other samples routinely tested in the laboratory.

Handling and safety requirements

Contact surfaces must be decontaminated with 70% alcohol or other suitable disinfectants. Used laboratory utensils must be cleaned and sterilized as per laboratory standard operating procedure. Unused materials and waste must be removed as per laboratory waste management protocol. This involves autoclaving and/or removal by an approved waste removal company.

Specific environmental conditions required for the participant to conduct tests

Virus isolation must be handled in a biological safety cabinet class 2+

Results recording

Only the supplied excel "Result entry forms" [R.E.F] should be used to capture the PT results. The Participants must fill in the cover page and answer all the questions before capturing the results on R.E.F.

Submission of PT results to JGK

All results must be submitted on or before the 25th of September 2017. Any results received after this date will not be included in the analysis. Virus isolation and PCR: submission date extended to 2nd of October 2017

Enquiries

Page 2 of 3

For any inquiry, please use the contact details provided above to contact JGK

Return of the proficiency test items

None

FD: freeze-dried