

Proficiency Test Instructions for Participants

21 AUGUST 2017 RUN

PURPOSE

To provide for detailed instructions for participants in compliance with the requirements of ISO 17043, clause 4.6.1

INSTRUCTIONS

Scheme reference	Samples			Samples reconstitution	Testing timeframe	Approved test methods	Sheet to be used for submission of results to JGK	Unit of measurement (Results)	Number of test replicates per sample
	Type	Identification (as per the labels)	Storage						
JGKPT02 Antimicrobial susceptibility	Animal sp.: <u>bovine</u> 2 FD cultures (mastitis pathogens)	ATB01M – ATB02M	2-8°C	Add 1 ml sterile DH ₂ O to the vial	Test immediately after reconstitution. Keep cool	Disc diffusion technique (Kirby Bauer)	R.E.F 1.2 (supplied)	Zone Ø in millimeters (express results: S, I, R) 'S': sensitive 'I': intermediate 'R': resistance	Culture: 1 rep. Susceptibility: Supplied discs: 3 repl. Own discs: 1 rep.
	Antimicrobial discs (x18)	Lincomycin (x6) Penicillin 10 µg (x6) Amikacin 30 µg (x6)		Discs are ready to use (Commercial discs)					
JGKPT26-28 Milk hygiene (TBC, Col, EC)	9 FD cultures	TBC01 – TBC03 COL01 – COL03 EC01 – EC03	2-8°C	Add 10 ml to the vials and dilute before testing as follows: TBC: 2 Dilutions COL: 2 dilutions EC: direct plating	Test immediately after reconstitution. Keep cool	3M Petri films	R.E.F 26-31 (supplied)	Colony counts	2 rep.
JGKPT29-31 Antimicrobial residues - Milk	4 FD milk	RES01M – RES04M	2-8°C	Add 3 ml sterile DH ₂ O to the vial Mix thoroughly	Test immediately. Keep cool	Rapid tests Snap Charm	R.E.F 26-31 (supplied)	Positive/Negative <u>Target drug</u>	1 rep.
JGKPT32-34 Antimicrobial residues - Meat	4 x 25 g meat	RES01F – RES04F	2-8°C	N/A Ready to process as per your SOP	Test immediately. Keep cool	Rapid and Chromatographic methods	R.E.F 32-44 (supplied)	Positive/Negative <u>Target drug</u>	1 rep.
JGKPT35 Food hygiene – C. jejuni	5 x 25 g meat	FH01 – FH05	2-8°C	N/A Ready to process as per your SOP	Test immediately. Keep cool	Standard veterinary microbiology ISO methods	R.E.F 32-44 (supplied)	Positive/Negative <u>Target bacteria</u>	1 rep.

JGKPT36 Food hygiene – Coliforms	2 FD cultures	COL01 – COL02	2-8°C	Add 1 ml sterile DH ₂ O to the vial	Test immediately after reconstitution. Keep cool	Standard veterinary microbiology ISO methods	R.E.F 32-44 (supplied)	Colony counts	2 rep.
JGKPT37 Food hygiene – <i>E. coli</i> O157	5 x 25 g meat	FH01 – FH05	2-8°C	N/A Ready to process as per your SOP	Test immediately. Keep cool	Standard veterinary microbiology ISO methods	R.E.F 32-44 (supplied)	Positive/Negative <u>Target bacteria</u>	1 rep.
JGKPT42 Food hygiene – <i>L. monocytogenes</i>	5 x 25 g meat	FH01 – FH05	2-8°C	N/A Ready to process as per your SOP	Test immediately. Keep cool	Standard veterinary microbiology ISO methods	R.E.F 32-44 (supplied)	Positive/Negative <u>Target bacteria</u>	1 rep.
JGKPT43 Food hygiene – Salmonella	5 x 25 g meat	FH01 – FH05	2-8°C	N/A Ready to process as per your SOP	Test immediately. Keep cool	Standard veterinary microbiology ISO methods	R.E.F 32-44 (supplied)	Positive/Negative <u>Target bacteria</u>	1 rep.
JGKPT44 Food hygiene – Total viable count	2 FD cultures	TVC01 – TVC02	2-8°C	Add 1 ml sterile DH ₂ O to the vial	Test immediately after reconstitution. Keep cool	Standard veterinary microbiology ISO methods	R.E.F 32-44 (supplied)	Colony counts	2 rep.
JGKPT45 Toxicology – Aflatoxin B1, B2, G1, G2 (maize)	Flower	MYC01 – MYC04 (S01 – S04)	Cool	N/A	When convenient Keep samples cool Observe deadline	HPLC ELISA	R.E.F 45-51 (supplied)	ppb	2 rep.
JGKPT51 Toxicology – Fumonisin B1, B2 (maize)	Flower	MYC01 – MYC04 Or S01 – S04	Cool	N/A	When convenient Keep samples cool Observe deadline	HPLC ELISA	R.E.F 45-51 (supplied)	ppb	2 rep.
JGKPT59 Somatic cell counting	6 x 20 ml milk (liquid) (preserved)	SCC01 – SCC06	2-8°C	N/A Ready to process as per your SOP	Test immediately. Keep cool	All counting devices	R.E.F 59-60 (supplied)	Cell counts	3 rep.
JGKPT60 Mastitis pathogens	2 x 10 ml milk (liquid)	MAST01 – MAST02	2-8°C	N/A Ready to process as per your SOP	Test immediately. Keep cool	Standard veterinary microbiology	R.E.F 59-60 (supplied)	Full ID <u>Target bacteria</u>	1 rep.

Treatment of samples

No special treatment should be given to the PT samples. They [samples] should be handled in the same manner as all the other samples routinely tested in the laboratory.

Handling and safety requirements

Contact surfaces must be decontaminated with 70% alcohol or other suitable disinfectants. Used laboratory utensils must be cleaned and sterilized as per laboratory standard operating procedure. Unused materials and waste must be removed as per laboratory waste management protocol. This involves autoclaving and/or removal by an approved waste removal company.

Specific environmental conditions required for the participant to conduct tests

None

Results recording

Only the supplied excel “Result entry forms” [R.E.F] should be used to capture the PT results. The Participants must fill in the cover page and answer all the questions before capturing the results on R.E.F.

Submission of PT results to JGK

All results must be submitted on or before the 11th of September 2017. **Any results received after this date will not be included in the analysis.**

Enquiries

For any inquiry, please use the contact details provided above to contact JGK

Return of the proficiency test items

None

FD: freeze-dried