

Proficiency Test Instructions for Participants

30 OCTOBER 2017 RUN

PURPOSE

To provide for detailed instructions for participants in compliance with the requirements of ISO 17043, clause 4.6.1

INSTRUCTIONS

Scheme reference	Samples			Samples reconstitution	Testing timeframe	Approved test methods	Sheet to be used for submission of results to JGK	Unit of measurement (Results)	Number of test replicates per sample
	Type	Identification (as per the labels)	Storage						
JGKPT01 Antimicrobial susceptibility	Animal sp.: <u>Poultry</u> 2 FD cultures (clinical pathogens)	ATB01C – ATB02C	2-8°C	Add 1 ml sterile DH ₂ O to the vial	Test immediately after reconstitution Observe deadline for submission of results	Disc diffusion technique (Kirby Bauer)	R.E.F 1.2 (supplied)	Zone Ø in millimeters (express results: S, I, R) 'S': sensitive 'I': intermediate 'R': resistance	Culture: 1 rep. Susceptibility: Supplied discs: 3 repl. Own discs: 1 rep.
	Antimicrobial discs (x18)	Doxycycline 30 µg (x6) Colistin sulphate 10 µg (x6) Tilmicosin 15 µg (x6)		Discs are ready to use (Commercial discs)					
JGKPT03 Brucella: RBT	1 FD bovine serum 1 liquid serum – Dilution serum	Sample 1: Batch 14423 (FD) Sample 2: Dilution serum	2-8°C	FD: Add 1000 µl sterile DH ₂ O to the FD serum Dilution serum is ready to use	Test within 5 days after reconstitution Observe deadline for submission of results	RBT	R.E.F 3.5 (supplied)	Positive/Negative	1 rep.
JGKPT04 Brucella: CFT				Read instructions carefully before testing – see below	Test within 5 days of dispatch Observe deadline for submission of results	CFT (DAFF approved SOP for SA Labs)	R.E.F 3.5 (supplied)	CFTIU/ml	2 rep.
JGKPT05 Brucella: MRT	4 fresh cow's milk	CAM01 – CAM04	2-8°C	N/A Ready to process	Preferably immediately but within 3 days from date of dispatch	MRT	R.E.F 3.5 (supplied)	Positive/Negative	1 rep.
JGKPT06 Brucella: Culture & Identification	3 FD cultures 1 Swab	2 FD sera: CAC01 – CAC02 1 FD milk – CAC03 1 Swab in trans – CAC04	2-8°C	Add 1 ml sterile DH ₂ O to FD sera Add 5 ml sterile DH₂O to FD milk	Test immediately after reconstitution Observe deadline for submission of results	Conventional microbiology and Molecular technique	R.E.F 06 (supplied)	Presence/Absence: <u>Brucella abortus</u> Strain: <u>Field or vaccine</u>	1 rep.

JGKPT07 Brucella: Slides stain & ID	4 unstained slides (4 replicates each)	Slide 1 – slide 4	Room T° (dust free)	N/A	Anytime, as long as deadline for submission of results is observed	Appropriate staining techniques	R.E.F 06 (supplied)	Positive/Negative <u>Brucella organisms</u>	1 rep.
JGKPT61-64 Trichomonas and Campylobacter: Culture/PCR	5 Cultures in transport media	SW01 – SW05	2-8°C	N/A Ready to process	Preferably immediately but no later than 5 days after dispatch	Conventional microbiology and Molecular technique	R.E.F 61.4 (supplied)	Presence/Absence: <u>Trichomonas foetus</u> , <u>Campylobacter fetus</u>	1 rep.

Treatment of samples

No special treatment should be given to the PT samples. They [samples] should be handled in the same manner as all the other samples routinely tested in the laboratory.

Handling and safety requirements

Contact surfaces must be decontaminated with 70% alcohol or other suitable disinfectants. Used laboratory utensils must be cleaned and sterilized as per laboratory standard operating procedure. Unused materials and waste must be removed as per laboratory waste management protocol. This involves autoclaving and/or removal by an approved waste removal company.

Specific environmental conditions required for the participant to conduct tests

JGKPT06 (Brucella Culture) must be handled in a biological safety cabinet class 2+

Results recording

Only the supplied excel “Result entry forms” [R.E.F] should be used to capture the PT results. The Participants must fill in the cover page and answer all the questions before capturing the results on R.E.F.

Submission of PT results to JGK

All results must be submitted on or before the **21st of November 2017**. **Any results received after this date will not be included in the analysis.**

Enquiries

For any inquiry, please use the contact details provided above to contact JGK

Return of the proficiency test items

None

FD: freeze-dried

Instruction for Brucella RBT and CFT

This time you will receive 2 samples only: 1 x FD serum (batch 14423) and 1 x Dilution serum (liquid)

1. Reconstitute FD serum by adding 1 ml sterile D.H₂O
2. The dilution serum is ready to use – add nothing
3. Set up a dilution series using Eppendorf tubes or any small tubes/containers. Mark them tube 1 to tube 7
4. Dispense 750 µl of the dilution serum (Point 2) into the 1st Eppendorf tube or any small tube/container
5. Dispense 500 µl of the dilution serum into Eppendorf tubes 2, 3, 4, 5 and 6. Leave tube 7 blank (add nothing)
6. Draw 250 µl of the reconstituted FD serum (Point 1) and transfer it to tube 1 to make up **1ml**
7. Using a new tip, mix well the content of the tube 1; then draw 500 µl and transfer to tube 2
8. Using a new tip, mix well the content of the tube 2; then draw 500 µl and transfer to tube 3
9. Using a new tip, mix well the content of tube 3; then draw 500 µl and transfer to tube 4
10. Use a new tip, mix well the content of tube 4; then draw 500 µl and transfer to tube 5

11. Use a new tip, mix well the content of tube 5; then draw 500 µl and transfer to tube 6.

Note: you now have **1 ml** in tube 6

12. Use a new tip; mix well the content of tube 6; then draw 500 µl and transfer to tube 7, which was left blank.

Note: tube 6 and tube 7 are identical replicates

Testing:

13. Perform RBT/CFT on the reconstituted FD serum and dilution serum as many times as you wish, but you will submit only 1 result for RBT and 2 results for CFT (not averaged results)
14. Perform RBT/CFT on each serial sample (Tube 1- tube 7) as many times as you wish, but you will submit only 1 result for RBT and 2 results for CFT (not averaged results)

Pipetting: for all your pipetting, go to the first stop of the plunger for drawing and the second stop for dispensing